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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁵: C12N 15/12, 15/63	A1	(11) International Publication Number: WO 94/02605 (43) International Publication Date: 3 February 1994 (03.02.94)
(21) International Application Number: PCT/US93/06782 (22) International Filing Date: 20 July 1993 (20.07.93) (30) Priority data: 918,314 22 July 1992 (22.07.92) US (71) Applicant: DUKE UNIVERSITY [US/US]; Erwin Road, Durham, NC 27741 (US). (72) Inventors: STILES, Gary, L.; 2463 Foxwood Drive, Chapel Hill, NC 27514 (US). REN, Hongzu; 200 Seven Oaks Road, Apt. 12B, Durham, NC 27704 (US). OLAH, Mark; 102 1/2 West Maynard Avenue, Durham, NC 27704 (US).		(74) Agents: SIBLEY, Kenneth, D. et al.; Bell, Seltzer, Park & Gibson, P.O. Drawer 34009, Charlotte, NC 28234 (US). (81) Designated States: AU, CA, JP, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i>
(54) Title: DNA ENCODING THE HUMAN A ₁ ADENOSINE RECEPTOR (57) Abstract Genomic and complementary DNA encoding the human A ₁ adenosine receptor are disclosed, along with vectors and host cells containing the same, oligonucleotides and antisense oligonucleotides directed to the same (particularly antisense oligonucleotides directed to an intron/exon junction), and methods of using the foregoing.		

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-1-

DNA ENCODING THE HUMAN A₁ ADENOSINE RECEPTOR

The present invention was made with Government support under grant number RO1 HL35134 from the National Heart Lung and Blood Institute. The Government has certain rights to this invention.

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Field of the Invention

The present invention relates to DNA encoding mammalian A₁ adenosine receptors, and particularly relates to intronless complimentary DNA encoding the human A₁ adenosine receptor and intron-containing genomic DNA encoding mammalian A₁ adenosine receptors.

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Background of the Invention

Adenosine is found in all living cells and can be released under appropriate conditions, such as ischemia or anoxia, where it can then act upon adenosine receptors to produce a variety of physiological effects. Adenosine receptors are now known to be integral membrane proteins which bind extracellular adenosine, thereby initiating a transmembrane signal via specific guanine nucleotide binding proteins known as G-proteins to modulate a variety of second messenger systems, including adenylyl cyclase, potassium channels, calcium channels and phospholipase C.

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-2-

See G. Stiles, *Clin. Res.* 38, 10-18 (1990); G. Stiles, *J. Biol. Chem.* 267, 6451-6454 (1992).

Adenosine receptors control a variety of important physiological effects including regulation of heart rate and contractility, regulation of smooth muscle tone in both blood vessels and the gastrointestinal tract, regulation of neurotransmitter release in brain, induction of sedation in the brain and regulation of platelet function. Although much biochemical and pharmacological information has become available on the two main types of adenosine receptors (known respectively as A₁ and A₂) which inhibit and stimulate adenylyl cyclase, much less information is available about their structure at the RNA and DNA level.

Adenosine receptors can be defined by an agonist potency series which, for the A₁ receptor, is R-PIA > NECA > S-PIA, and which for the A₂ receptor is NECA > R-PIA > S-PIA. See R. Olsson and J. Pearson, *Physiol. Rev.* 70, 761-845 (1990). Very recently we have found evidence for a unique A₁ adenosine receptor which is expressed in the bovine brain which has a different potency series such that R-PIA is > S-PIA which is > NECA. We have cloned and sequenced this receptor and have begun studies on site-directed mutagenesis to understand the ligand binding site. See M. Olah et al., *J. Biol. Chem.* 267, 10764-10770 (1992). In addition, we have found a new receptor previously not suspected, which we have termed the A₃ adenosine receptor. This receptor has likewise been cloned, sequenced and expressed. See F. Zhou et al., *Proc. Natl. Acad. Sci. USA*, in press (1992).

Summary of the Invention

To date, there has been no information on the genomic structure of any adenosine receptor. Neither has there been any information available on any human adenosine receptor cDNA. Such information is, however, necessary if

-3-

the genomic structure and activity of the human receptor is to be explored. This information is provided herein.

A first aspect of the present invention is isolated DNA encoding an A₁ adenosine receptor selected from the group consisting of: (a) isolated DNA which encodes the human genomic A₁ adenosine receptor of SEQ ID NO:6, which isolated DNA contains the DNA sequences given herein as SEQ ID NO:1 and SEQ ID NO:3; (b) isolated human genomic DNA which hybridizes to isolated DNA of (a) above under conditions represented by a wash stringency of 0.3M NaCl, 0.03M sodium citrate, and 0.1% SDS at 60°C, which is at least 65% homologous to the isolated DNA of (a) above (i.e., 65% homologous or more with respect to total DNA sequence; homology with respect to exon sequence alone is about 93% or more), and which encodes a human A₁ adenosine receptor; and (c) isolated DNA differing from the isolated DNAs of (a) and (b) above in nucleotide sequence due to the degeneracy of the genetic code, and which encodes a human A₁ adenosine receptor.

A second aspect of the present invention is isolated DNA encoding an A₁ adenosine receptor selected from the group consisting of: (a) the isolated DNA which encodes a human A₁ adenosine receptor and has the DNA sequence given herein as SEQ ID NO:5; (b) isolated DNA which hybridizes to the isolated DNA of (a) above under conditions represented by a wash stringency of 0.3M NaCl, 0.03M sodium citrate, and 0.1% SDS at 60°C, which is at least 93% homologous to isolated DNA of (a) above, and which encodes a human A₁ adenosine receptor; and (c) isolated DNA differing from the isolated DNAs of (a) and (b) above in nucleotide sequence due to the degeneracy of the genetic code, and which encodes an A₁ adenosine receptor.

A third aspect of the present invention is a recombinant DNA sequence comprising vector DNA and a DNA as given above which encodes an A₁ adenosine receptor.

A fourth aspect of the present invention is a host cell containing a recombinant DNA sequence as given

above and capable of expressing the encoded A₁ adenosine receptor.

A fifth aspect of the present invention is an oligonucleotide (e.g., a polynucleotide 10-200 nucleotides in length, preferably 20-200 nucleotides in length) capable of selectively hybridizing to DNA which comprises a portion of a gene coding for an A₁-adenosine receptor as given herein. Such oligonucleotides may comprise fragments of a DNA as given above which encodes an A₁ adenosine receptor. Such oligonucleotides may be used in a suitable vector for carrying out homologous recombination. When labelled with a detectable group, such oligonucleotides comprise probes. In one preferred embodiment, such oligonucleotides comprise the introns, fragments of the introns, or hybridize to the introns of, the genomic A₁-adenosine receptor DNA disclosed herein (i.e., from 5' to 3', the first, second, third, fourth or fifth intron of the genomic A₁-adenosine receptor) which selectively hybridize to the introns of a genomic A₁-adenosine receptor DNA.

A sixth aspect of the present invention is isolated DNA as given above, and oligonucleotides as given above, configured in antisense for the production of antisense RNA which would interfere with the expression of native A₁-adenosine receptor (preferably selected so as to not interfere with the production of other adenosine receptors such as the A₂ or A₃ adenosine receptor (hereinafter referred to as "antisense DNAs"). Such antisense DNAs may be provided in a vector as given above for transcription in a suitable cell where they then interfere with the production of the A₁ adenosine receptor. Preferably, such antisense DNAs are directed to one of the intron-exon junctions of the A₁ adenosine receptor genomic DNA, as described herein.

A seventh aspect of the present invention is isolated DNA encoding mutant A₁ adenosine receptors in which (a) the receptors have decreased affinity for A₁ adenosine receptor agonists and/or antagonists, or (b) the receptors

-5-

bind A₁ adenosine receptor agonists with high affinity but fail to inhibit adenylyl cyclase in cells transfected therewith (in which cells the receptor would, but for the mutation, otherwise be seen inhibit adenylyl cyclase). For the latter, mutations which render the hydrophobic pocket in the fifth transmembrane domain less hydrophobic are particularly preferred.

The DNA sequence information provided herein is valuable for several reasons. First, it is known that it regulates a wide variety of physiological effects and, therefore, has great potential for use as a therapeutic agent in a variety of conditions ranging from manipulation of cardiac function, protection against ischemia, regulation of smooth muscle tone in blood vessels, potential usefulness as an agent in cystic fibrosis, usefulness as a potential agent in treatment of seizure activity. Recent studies have shown that the endogenous release of adenosine during a brief occlusion of a coronary artery leads to the protection of the myocardium from subsequent prolonged ischemic and anoxic events. This work shows that this appears to be mediated specifically via the A₁ adenosine receptor. See G. Liu et al., *Circulation* 84, 350-356 (1991); S. Ely and R. Berne, *Circulation* 85, 893-904 (1992). This indicates that manipulation of A₁ adenosine receptors through either specific ligand manipulation or through introduction by recombinant DNA techniques of wild-type or mutant receptors into myocardium may provide a protective effect against ischemia.

The foregoing and other aspects of the present invention are explained in detail in the drawings, Examples, and Detailed Description set forth below.

Brief Description of the Drawings

Figure 1 provides a schematic representation of the human A₁ adenosine receptor gene. Shown are the known structures of the exons indicated by E1, E2, etc., and the introns indicated by capital I. Each exon displays the

type of coding structure contained within its domain. For example, E1, E2 and E3 solely code for 5' untranslated regions of the messenger RNA. E4 codes for part of the 5' untranslated and the initial coding sequence of the receptor. E5 codes for a short stretch of coding region and E6 codes from the fourth transmembrane domain to the end of the coding sequence as well as the 3' untranslated region. Introns range in size from 300 b to greater than 10 kb.

Figure 2 shows the predicted structure of the human A₁ adenosine receptor as predicted by hydropathy plots. Demonstrated are the potential phosphorylation sites for protein kinase C and protein kinase A. In addition, potential glycosylation sites are shown by the asterisk or the small "fork-like" structure demonstrated coming off the second extracellular loop at the top of the figure. In addition are shown the potential site for fatty acid acylation on the carboxy terminal tail.

Detailed Description of the Invention

Amino acid sequences disclosed herein are presented in the amino to carboxy direction, from left to right. The amino and carboxy groups are not presented in the sequence. Nucleotide sequences are presented herein by single strand only, in the 5' to 3' direction, from left to right. Nucleotides and amino acids are represented herein in the manner recommended by the IUPAC-IUB Biochemical Nomenclature Commission, or (for amino acids) by three letter code, in accordance with 37 CFR §1.822 and established usage. See, e.g., PatentIn User Manual, 99-102 (Nov. 1990) (U.S. Patent and Trademark Office, Office of the Assistant Commissioner for Patents, Washington, D.C. 20231); U.S. Patent No. 4,871,670 to Hudson et al. at Col. 3 lines 20-43 (applicants specifically intend that the disclosure of this and all other patent references cited herein be incorporated herein by reference).

A. DNAs ENCODING A₁-ADENOSINE RECEPTORS AND OLIGONUCLEOTIDES THEREOF

DNAs which encode A₁-adenosine receptors, whether they are cDNAs or intron-containing genomic DNAs, encode a protein which, on expression in a suitable host cell, (a) selectively and stereospecifically binds adenosine, and (b) inhibits adenylate cyclase activity upon binding adenosine. This definition is intended to encompass natural allelic variations in the DNAs. Genomic DNAs of the present invention may code for an A₁-adenosine receptor of any species of origin, including mouse, rat, rabbit, cat, porcine, and human, but preferably code for an A₁-adenosine receptor of mammalian origin, and most preferably code for a human A₁-adenosine receptor. Complementary DNAs of the present invention encode human A₁-adenosine receptors. Hybridization conditions which will permit other DNA sequences which code on expression for an A₁-adenosine receptor to hybridize to a DNA sequence as given herein are, in general, high stringency conditions. For example, hybridization of such sequences may be carried out under conditions represented by a wash stringency of 0.3 M NaCl, 0.03 M sodium citrate, 0.1% SDS at 60°C or even 70°C to DNA disclosed herein in a standard in situ hybridization assay. See J. Sambrook et al., Molecular Cloning, A Laboratory Manual (2d Ed. 1989) (Cold Spring Harbor Laboratory)). The same hybridization conditions are used to determine hybridization of oligonucleotides. In general, genomic DNA sequences which code for A₁-adenosine receptors and hybridize to the genomic DNA sequence encoding the human A₁-adenosine receptor disclosed herein will be at least 65%, 70%, 75%, 80%, 85%, 90%, or even 95% homologous or more with the sequence of the genomic DNA encoding the human A₁-adenosine receptor disclosed herein (with respect to the total genomic DNA). These same levels of homology apply to oligonucleotide probes which hybridize to the introns of the genomic DNA sequences disclosed herein. Homology among the exons alone between various human genomic DNA sequences

encoding the A₁ adenosine receptor are contemplated to be of the same order as given below with respect to cDNA sequences.

In general, complementary DNA sequences which
5 encode human A₁-adenosine receptors which hybridize the the
cDNA encoding the human A₁-adenosine receptor disclosed
herein will be 93%, 94%, 95%, 96%, or even 97% homologous
or more to the cDNA sequence encoding the human A₁-adenosine
receptor disclosed herein. These same levels of homology
10 apply to oligonucleotides which hybridize to the human A₁-
adenosine receptor cDNA or gDNA disclosed herein.

Further, DNA sequences (or oligonucleotides)
which code for the same A₁-adenosine receptor (or fragment
thereof) as coded for by the foregoing sequences, but which
15 differ in codon sequence from these due to the degeneracy
of the genetic code, are also an aspect of this invention.
The degeneracy of the genetic code, which allows different
nucleic acid sequences to code for the same protein or
peptide, is well known in the literature. See e.g., U.S.
20 Patent No. 4,757,006 to Toole et al. at Col. 2, Table 1.

B. GENETIC ENGINEERING TECHNIQUES

The production of cloned genes, recombinant DNA,
vectors, transformed host cells, proteins and protein
fragments by genetic engineering is well known. See, e.g.,
25 U.S. Patent No. 4,761,371 to Bell et al. at Col. 6 line 3
to Col. 9 line 65; U.S. Patent No. 4,877,729 to Clark et
al. at Col. 4 line 38 to Col. 7 line 6; U.S. Patent No.
4,912,038 to Schilling at Col. 3 line 26 to Col. 14 line
12; and U.S. Patent No. 4,879,224 to Wallner at Col. 6 line
30 8 to Col. 8 line 59.

A vector is a replicable DNA construct. Vectors
are used herein either to amplify DNA encoding A₁-adenosine
receptors as given herein and/or to express DNA which
encodes A₁-adenosine receptors as given herein. An
35 expression vector is a replicable DNA construct in which a
DNA sequence encoding an A₁-adenosine receptor is operably

linked to suitable control sequences capable of effecting the expression of the receptor in a suitable host. The need for such control sequences will vary depending upon the host selected and the transformation method chosen.

5 Generally, control sequences include a transcriptional promoter, an optional operator sequence to control transcription, a sequence encoding suitable mRNA ribosomal binding sites, and sequences which control the termination of transcription and translation.

10 Amplification vectors do not require expression control domains. All that is needed is the ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants.

15 Vectors comprise plasmids, viruses (e.g., adenovirus, cytomegalovirus), phage, and integratable DNA fragments (i.e., fragments integratable into the host genome by recombination). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself.

20 Expression vectors should contain a promoter and RNA binding sites which are operably linked to the gene to be expressed and are operable in the host organism.

25 DNA regions are operably linked or operably associated when they are functionally related to each other. For example, a promoter is operably linked to a coding sequence if it controls the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to permit translation.

30 Transformed host cells are cells which have been transformed or transfected with vectors containing a DNA sequence as disclosed herein constructed using recombinant DNA techniques. Transformed host cells ordinarily express the receptor, but host cells transformed for purposes of
35 cloning or amplifying the receptor DNA do not need to express the receptor.

-10-

Suitable host cells include prokaryote, yeast or higher eukaryotic cells such as mammalian cells and insect cells. Cells derived from multicellular organisms are a particularly suitable host for recombinant A₁-adenosine receptor synthesis, and mammalian cells are particularly preferred. Propagation of such cells in cell culture has become a routine procedure (Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)). Examples of useful host cell lines are VERO and HeLa cells, Chinese hamster ovary (CHO) cell lines, and WI138, BHK, COS-7, CV, and MDCK cell lines. Expression vectors for such cells ordinarily include (if necessary) an origin of replication, a promoter located upstream from the DNA encoding the A₁-adenosine receptor to be expressed and operatively associated therewith, along with a ribosome binding site, an RNA splice site (if intron-containing genomic DNA is used), a polyadenylation site, and a transcriptional termination sequence.

The transcriptional and translational control sequences in expression vectors to be used in transforming vertebrate cells are often provided by viral sources. For example, commonly used promoters are derived from polyoma, Adenovirus 2, and Simian Virus 40 (SV40). See, e.g., U.S. Patent No. 4,599,308.

An origin of replication may be provided either by construction of the vector to include an exogenous origin, such as may be derived from SV 40 or other viral (e.g. Polyoma, Adenovirus, VSV, or BPV) source, or may be provided by the host cell chromosomal replication mechanism. If the vector is integrated into the host cell chromosome, the latter is often sufficient. Rather than using vectors which contain viral origins of replication, one can transform mammalian cells by the method of cotransformation with a selectable marker and the receptor DNA. Examples of suitable selectable markers are dihydrofolate reductase (DHFR) or thymidine kinase. This method is further described in U.S. Pat. No. 4,399,216.

-11-

Other methods suitable for adaptation to the synthesis of the A₁-adenosine receptor in recombinant vertebrate cell culture include those described in M-J. Gething et al., Nature 293, 620 (1981); N. Mantei et al., Nature 281, 40; A. Levinson et al., EPO Application Nos. 117,060A and 117,058A.

Host cells such as insect cells (e.g., cultured *Spodoptera frugiperda* cells) and expression vectors such as the baculovirus expression vector (e.g., vectors derived from *Autographa californica* MNPV, *Trichoplusia ni* MNPV, *Rachiplusia ou* MNPV, or *Galleria ou* MNPV) may be employed in carrying out the present invention, as described in U.S. Patents Nos. 4,745,051 and 4,879,236 to Smith et al. In general, a baculovirus expression vector comprises a baculovirus genome containing the gene to be expressed inserted into the polyhedrin gene at a position ranging from the polyhedrin transcriptional start signal to the ATG start site and under the transcriptional control of a baculovirus polyhedrin promoter.

Prokaryote host cells include gram negative or gram positive organisms, for example *Escherichia coli* (E. coli) or *Bacilli*. Higher eukaryotic cells include established cell lines of mammalian origin as described below. Exemplary host cells are E. coli W3110 (ATCC 27,325), E. coli B, E. coli X1776 (ATCC 31,537), E. coli 294 (ATCC 31,446). A broad variety of suitable prokaryotic and microbial vectors are available. E. coli is typically transformed using pBR322. Promoters most commonly used in recombinant microbial expression vectors include the beta-lactamase (penicillinase) and lactose promoter systems (Chang et al., Nature 275, 615 (1978); and Goeddel et al., Nature 281, 544 (1979)), a tryptophan (trp) promoter system (Goeddel et al., Nucleic Acids Res. 8, 4057 (1980) and EPO App. Publ. No. 36,776) and the tac promoter (H. De Boer et al., Proc. Natl. Acad. Sci. USA 80, 21 (1983)). The promoter and Shine-Dalgarno sequence (for prokaryotic host expression) are operably linked to the DNA encoding the A₁-

-12-

adenosine receptor, i.e., they are positioned so as to promote transcription of A₁-adenosine receptor messenger RNA from the DNA.

Eukaryotic microbes such as yeast cultures may also be transformed with vectors carrying the isolated DNA's disclosed herein. see, e.g., U.S. Patent No. 4,745,057. *Saccharomyces cerevisiae* is the most commonly used among lower eukaryotic host microorganisms, although a number of other strains are commonly available. Yeast vectors may contain an origin of replication from the 2 micron yeast plasmid or an autonomously replicating sequence (ARS), a promoter, DNA encoding the receptor as given herein, sequences for polyadenylation and transcription termination, and a selection gene. An exemplary plasmid is YRp7, (Stinchcomb et al., Nature **282**, 39 (1979); Kingsman et al., Gene **7**, 141 (1979); Tschemper et al., Gene **10**, 157 (1980)). Suitable promoting sequences in yeast vectors include the promoters for metallothionein, 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem. **255**, 2073 (1980) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Reg. **7**, 149 (1968); and Holland et al., Biochemistry **17**, 4900 (1978)). Suitable vectors and promoters for use in yeast expression are further described in R. Hitzeman et al., EPO Publ. No. 73,657.

C. USES OF GENOMIC AND COMPLEMENTARY DNAS ENCODING A₁-ADENOSINE RECEPTORS

1. In general. A₁-adenosine receptors made from cloned genes in accordance with the present invention may be used for screening compounds for A₁-adenosine receptor activity, or for determining the amount of a adenosine receptor agonist or antagonist in a solution (e.g., blood plasma or serum). For example, host cells may be transformed with a vector of the present invention, A₁-adenosine receptors expressed in that host, the cells lysed, and the membranes from those cells used to screen compounds for A₁-adenosine receptor binding activity.

-13-

Competitive binding assays in which such procedures may be carried out are well known. By selection of host cells which do not ordinarily express an adenosine receptor, preparations free of other receptors which might interfere with the assay, such as A_2 -adenosine receptors, can be obtained. Further, A_1 -adenosine receptor agonists and antagonists can be identified by transforming host cells with vectors of the present invention, which host cells also express adenylyl cyclase. Membranes obtained from such cells can be used in binding studies wherein the activity of the adenylyl cyclase is monitored. A_1 -adenosine receptor agonists will inhibit the adenylyl cyclase. Such cells must be capable of operatively associating the A_1 -adenosine receptor with the adenylyl cyclase, i.e., G protein must also be present in the cell membranes in the appropriate configuration. Thus, a further aspect of the present invention is an aqueous solution containing cell membranes, the cell membranes containing an adenosine receptor and adenylyl cyclase, wherein the cell membranes are essentially free of A_2 -adenosine receptors, and wherein the A_1 -adenosine receptors are capable of inhibiting the adenylyl cyclase on binding an A_1 -adenosine receptor agonist. A still further aspect of the present invention is an assay procedure comprising the steps of: (a) providing an aqueous solution containing cell membranes as described above; then (b) adding a test compound to the aqueous solution; and then (c) monitoring the activity of adenylyl cyclase in the aqueous solution.

DNAs of the present invention are useful in gene therapy, as discussed in greater detail below. For such purposes, retroviral vectors as described in U.S. Patent No. 4,650,764 to Temin and Watanabe or U.S. Patent No. 4,861,719 to Miller may be employed. DNAs of the present invention, or fragments thereof, may also be used in gene therapy carried out by homologous recombination or site-directed mutagenesis. See generally Thomas, K. and Capecchi, M., Cell 51, 503-512 (1987); Bertling, W.,

Bioscience Reports 7, 107-112 (1987); Smithies, O. et al., Nature 317, 230-234 (1985).

2. Regulation of blood vessel tone and modulation of restenosis following vessel manipulation.

5 Adenosine receptors both A₁ and A₂ subtypes are known to exist on both vascular smooth muscle cells and endothelial cells (Mol. Pharmacol. 37:149, 1990). These receptors are known to produce vasodilatation through activation of both
10 adenylyl cyclase (cAMP) and guanylate cyclase (cGMP) (Physiol. Rev. 70:761, 1990). The endogenous substance adenosine is responsible for producing vasodilatation in all vascular beds except renal and perhaps pulmonary, where constriction occurs (Physiol. Rev. 70:761, 1990; Circ. 65:1516, 1989). In addition, vascular constriction can be
15 overcome by adenosine's action. Recombinant DNA constructs such as the vectors described herein can be introduced into the blood vessel via catheter based techniques (Circ. 83:2007, 1991) to permit the transfection of the vessel with exogenous gene products to overexpress or inhibit the
20 activity of the A₁ adenosine receptor.

Thus, a further aspect of the present invention is a method of treating a human or animal (e.g., horse, cow) subject by administering a vector carrying a DNA
25 sequence or oligonucleotide or fragment thereof as described herein into a blood vessel of the subject (i.e., artery, vein) in an amount sufficient so that (depending on the nature of the vector and the specific purpose as discussed below) expression of the A₁ adenosine receptor is increased or inhibited.

30 For example, a restriction fragment of the human A₁AR cDNA could be inserted into pCMV or retroviral vectors. Retroviral vectors have certain advantages in that they have high efficiency for transfection and are effective in a wide range of cell types. The disadvantages are their
35 potential for inducing malignant transformation or producing a competent virus and hence produce an infection. These techniques are discussed in Wilson et al., Human Gene

Therapy 3, 179 (1991), which describes the transformation of human liver cells deficient in LDL receptors with DNA encoding LDL receptors using retroviral vectors. Similar approaches with DNAs encoding adenosine receptors could be accomplished.

A specific example is as follows. A cDNA as given herein is cloned into the MuLV vector which is a MO-MuLV derived vector containing ψ packaging sequences, transposon TN5 gene which encodes neomycin phosphotransferase, SV40 origin of replication, pBR origin of replication: beta actin promoter and unique Bam H1 cloning site. Once constructed the plasmids are isolated, purified and analyzed by restriction digestion using standard procedures.

Plasmid constructs are transfected with 3T3 ψ 2 packaging cell line and media from these cells can be used to infect the 3T3 ψ AM packaging cell line. G418 resistant clones of 3T3 ψ AM cells are isolated and used as a source of retroviruses for infecting target cells (J. Virol. 31:360, 1990; Cell 33:153, 1983; Proc. Natl. Acad. Sci. 31:6349, 1984). This technique could of course be modified in a number of ways. For example, the more recently described ψ CRIP packaging cell line could also be utilized (Proc. Natl. Acad. Sci. USA 85:6460, 1988). Once the vectors are made they can be mixed with a lipofectin mixture as described (Circ. 83:2007, 1991) and then introduced in a blood vessel using a perfusion balloon catheter in accordance with known techniques. Recent data suggest that lipofectin may not be necessary. In this manner A₁AR can be overexpressed to enhance the activity of A₁AR to module vascular tone/growth properties. Similar receptors in which sites for phosphorylation (J. Biol. Chem. 264:12657, 1989; Mol. Pharmacol. 40:639, 1991), which lead to desensitization of the receptor, are mutated out could be introduced to thwart the process of desensitization. Although two expression vectors are described above, the A₁AR could also be expressed using a wide range of

-16-

expression vectors including those with regulatable promoters either with heavy metal promoters or glucocorticoid promoters, etc. In addition, transfection systems utilizing site specific cogeners, such as
5 antibody/lipid mixtures or DNA constructs attached to antibodies for targeting, could also be used.

The receptors to be transfected into vessels would not be limited to wild type human A_1AR but mutants which are constitutively active or inactive, receptors
10 which couple to G protein other than G_i (such as G_s or G_o) constructed by interchanging the third cytoplasmic loop from receptors such as the β -adrenergic or A_2 adenosine receptors which couple to G_s for the third cytoplasmic loop of A_1AR .

15 In addition, one can express antisense RNA from the human A_1 -adenosine receptor DNAs disclosed herein to block production of the A_1AR , particularly antisense RNA directed to one of the intron-exon junctions of the genomic A_1 sequence.

20 Another potential use is to introduce these wild type or mutant receptors at the time of coronary angioplasty to modulate the vessels' reactivity at the site of balloon injury. These transfections could be used to promote enhanced vasodilation or to change the growth
25 properties of the endothelial cells or smooth muscle cells. Further, adenosine has been reported to enhance collateral formation and angiogenesis. Thus, by introducing receptor constructs one could modulate growth of vessels in the heart or other organs.

30 3. Cardio-protective effects. As described above, there is now evidence that protection against myocardial infarction afforded by the preconditioning, i.e., short-term occlusion of a coronary artery is mediated by A_1 adenosine receptors (Circ. 84:350, 1991). It appears
35 that activation of the A_1 adenosine receptor by adenosine during a brief ischemic event by some mechanism as yet unclear protects the heart to subsequent ischemic and

-17-

anoxic conditions. Therefore, enhancement of the effects of A₁ adenosine receptors may lead to greater protection of the myocardium to ischemia. This would have direct impact during procedures such as coronary artery bypass grafting, during coronary angioplasty, or in patients at high risk for myocardial infarction.

Thus, an additional aspect of the present invention is a method of combatting ischemia in a human or animal subject in need of such treatment, comprising administering to the subject (e.g., intravenously) a vector carrying a DNA as given herein, and capable of expressing the same in myocardial tissue, in an amount effective to combat ischemia in said subject.

There are several approaches that could be taken to implement enhanced A₁ receptor function. One would be to over-express wild type human A₁ receptors in the myocardium so that, for a given level of adenosine present extracellularly, there would be an enhanced signal through the G-protein mechanism to create an enhanced protective effects. This could be accomplished by inserting the cDNA fragments, as described above, into an expression vector which could be targeted for myocardial cells based on specific antigens on the surface of cardiac cells. A second approach is that, by understanding the promoter/enhancer region of the gene, one can specifically target means of activating a promoter/enhancer region to increase the transcription and ultimately translation of the messenger RNA for the A₁ receptor to enhance the relative levels of A₁ receptors in the heart. Multiple approaches are currently being studied worldwide (Trends in Cardio. Med. 7:271, 1991).

4. Control of arrhythmias. Adenosine has been found to be a very effective antiarrhythmic agent for supraventricular tachycardias (Circ. 83:1499, 1991). The effects are known to occur through its action on A₁ adenosine receptors on the SA node and the AV node. The arteries subserving these regions of the heart are now

-18-

accessible through cardiac catheterization techniques and, therefore, direct access to these areas can be obtained with the potential for delivering specific vectors containing cDNA sequences of the A₁ adenosine receptor.

5 There are at least two potential uses, one for the treatment of supraventricular tachycardias in which additional wild-type A₁ receptors could be delivered to either the AV node or the SA node, thereby increasing the effectiveness of endogenous adenosine on slowing the SA and

10 AV node. Another potential treatment is for bradyarrhythmias in which there is excessive slowing of the SA and AV node, in which case antisense RNA could be expressed in this region by the introduction of a vector carrying the appropriate cDNA to block the production of

15 the A₁ receptor in these areas, thus diminishing the effect of endogenous adenosine on heart rate and allowing an increased heart rate to occur. For all the above applications, one would utilize transient transfections likely working through an episomal mode since these cells

20 do not replicate and, therefore, stable transfections are not likely to occur.

5. CNS therapies. The same type of protective effect as described in the heart may well also occur in the brain during strokes. Therefore, there is the potential

25 for delivering the same type of constructs described for the cardioprotective effect above into regions of the brain subject to ischemia.

6. Treatment of cystic fibrosis. There has been recent evidence published whereby blockade of the A₁ adenosine receptors from cystic fibrosis cells activates chloride efflux from these cells. This work was able to document that the A₁ adenosine receptor antagonist known as cyclopentyl-1,3-dipropylxanthine activated chloride efflux from a cell line derived from cystic fibrosis patients

30 which is known to contain mutant phenylalanine 508 characteristic of cystic fibrosis. There would be several potential mechanisms for blocking the effects of endogenous

35

-19-

adenosine in these cells. First, one could transfect into these cells DNA encoding a human A_1 -adenosine receptor which would bind adenosine but would not activate the effector system. Examples of these approaches include the construction of an adenovirus vector which is a replication deficient recombinant adenovirus such as the Ad-CITR construct used in cystic fibrosis. This is constructed from the adenovirus type 5 (Ad5) deletion mutant, Ad-d1324 and a plasmid (pTG 595T) as described (Cell 68:143, 1992). This or other vectors could be delivered in an aerosolized form in lipid or saline or could be introduced with cell specificity using an ectopic murine retrovirus to infect only the chosen cell through antibody mediated binding of a cell surface receptor. Alternatively, polycations such as poly-L-lysine to encapsulate and deliver the construct to the cell surface (Thorax. 46:46, 1991) thus creating a receptor which was constitutently blocked. Another mechanism would be to use antisense approaches to block synthesis of the receptor as described above. Additionally, one could use pharmacological methods (covalent ligands) to block receptors with agents that would specifically bind to the receptor and covalently inhibit its activity.

7. Ocular uses of DNA encoding A_1 AR. Recent information (Current Eye Research 11:453, 1992) demonstrates that R-PIA (an A_1 AR selective ligand) at selective doses when applied to the eye of rabbits leads to a significant reduction in intraocular pressure (5-8 mmHg) and this response is blocked by adenosine receptor antagonist. These data open the way for introducing constitutively active A_1 AR, A_1 AR resistant to desensitization or over expression of wild type A_1 AR into interior chamber structures such as the ciliary body, muscles, etc., in order to enhance A_1 AR efforts and lower extraocular pressure. The same type of vectors and constructs as described above could be used here. Likely the constructs would need to be injected for delivery.

-20-

8. Antisense oligonucleotides. Another aspect of this invention is an antisense oligonucleotide having a sequence capable of binding specifically with any sequences of an mRNA molecule which encodes a human A₁ adenosine receptor so as to prevent translation of the mRNA molecule (binding conditions may be at the stringencies as given above with respect to DNA hybridization). The antisense oligonucleotide may have a sequence capable of binding specifically with any sequences of the DNA molecule whose sequence is disclosed herein in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5. Chemical analogs of nucleotides (e.g., nucleotides in which the phosphodiester bonds have been modified, e.g., to the methylphosphonate, the phosphotriester, the phosphorothioate, the phosphorodithioate, or the phosphoramidate, so as to render the oligonucleotide more stable *in vivo*) are specific examples of such antisense oligonucleotides. Antisense oligonucleotides may be of any suitable length (e.g., from about 10 to 60 nucleotides in length), depending on the particular target being bound and the mode of delivery thereof. Preferably the antisense oligonucleotide is directed to an mRNA region containing a junction between intron and exon. Where the antisense oligonucleotide is directed to an intron/exon junction, it may either entirely overlie the junction or may be sufficiently close to the junction to inhibit splicing out of the intervening exon during processing of precursor mRNA to mature mRNA (e.g., with the 3' or 5' terminus of the antisense oligonucleotide being is positioned within about, for example, 10, 5, 3, or 2 nucleotides of the intron/exon junction).

Pharmaceutical compositions comprising an antisense oligonucleotide as given above effective to reduce expression of a human A₁ adenosine receptor by passing through a cell membrane and binding specifically with mRNA encoding a human A₁ adenosine receptor in the cell so as to prevent its translation are another aspect of the present invention. Such compositions are provided in a

suitable pharmaceutically acceptable carrier (e.g., sterile pyrogen-free saline solution). The antisense oligonucleotides may be formulated with a hydrophobic carrier capable of passing through a cell membrane (e.g., in a liposome, with the liposomes carried in a pharmaceutically acceptable aqueous carrier). The oligonucleotides may also be coupled to a substance which inactivates mRNA, such as a ribozyme. Such oligonucleotides may be administered to a subject to inhibit the activation of A₁ adenosine receptors, which subject is in need of such treatment for any of the reasons discussed herein.

9. Additional uses. DNAs of the present invention, and oligonucleotides derived therefrom, are useful for screening for restriction fragment length polymorphism (RFLP) associated with disorders such as cystic fibrosis or other disorders potentially involving a defective A₁-adenosine receptor (or defective regulation thereof).

Oligonucleotides of the present invention are useful as diagnostic tools for probing A₁-adenosine receptor expression in various tissues. For example, tissue can be probed in situ with oligonucleotide probes carrying detectable groups (e.g., a radioisotope such as ³²P, ¹²⁵I, ¹³¹I, ³H, ¹⁴C, or ³⁵S; an enzyme such as horseradish peroxidase, or alkaline phosphatase; an electron dense ligand such as ferritin or gold) by conventional autoradiography techniques to investigate native expression of this receptor or pathological conditions relating thereto. Further, chromosomes can be probed to investigate the presence or absence of an A₁-adenosine receptor gene, and potential pathological conditions related thereto.

The present invention is explained in greater detail in the following Examples. These Examples are for illustrative purposes only, and are not to be taken as limiting of the invention.

-22-

EXAMPLES

I. METHODS

[γ - 32 P]ATP, [35 S]dATP, [α - 32 P]ATP and [3 H]XAC were from DuPont-New England Nuclear. Restriction enzymes and T4 DNA ligase were from either Boehringer-Mannheim or Promega. The pCMV5 expression vector was obtained from Dr. Marc Caron (Duke University Medical Center) and was originally described by Dr. David Russell (University of Texas). All cell culture supplies were from Gibco.

10 Genomic Cloning. The human A₁ adenosine receptor (A₁AR) genomic clones were isolated from human leukocyte genomic library (Clontech) in EMBL3 with various probes. In general, each time $\sim 1.0 \times 10^6$ plaques were screened. Duplicate nylon filters (Biotrans, ICN) with lifted plaques were prehybridized in 50% formamide (30% with shorter probe), 5x SSPE (1x SSPE = 0.15 M NaCl, 0.01 M Na₂HPO₄ and 1 mM EDTA, pH 7.4), 5x Denhardt's solution, 0.1% SDS and 0.1 mg/ml salmon sperm DNA for 4 hrs at 42°C. Hybridization was conducted in the same solution without Denhardt's solution, plus radiolabeled probe ($\sim 500,000$ cpm/mo) for about 16 hrs at 42°C. The filters were then washed twice in 2x SSC (1x SSC = 0.15 M NaCl and 0.015 M sodium citrate) and 0.1% SDS at room temperature for 15 min each. The final wash conditions were determined according to the length of probe used for screening, ranging from 1x SSC, 0.1% SDS, 55°C and 10 min to 0.1x SSC, 0.1% SDS, 60°C and 10 min.

30 The structure of the human genomic adenosine A₁ receptor gene is set forth in SEQ ID NO:1 and SEQ ID NO:3, with the amino acid fragment coded in SEQ ID NO:1 set forth in SEQ ID NO:2, and with the amino acid fragment coded in SEQ ID NO:3 set forth in SEQ ID NO:4. Note that SEQ ID NO:1 contains part of intron 1, all of exon 2, all of intron 3, all of exon 4, and part of intron 4, while SEQ ID NO: 3 contains part of intron 5, and all of exon 6. Portions not shown can be derived from the portions shown by standard techniques such as polymerase chain reaction

-23-

(PCR) or other of the known amplification techniques, taken with the information provided in all of the sequences herein. For example, it can be determined from these materials that exon number 5 contains nucleotides 752 to 804 of SEQ ID NO:5, and the introns immediately flanking exon number 5 can be produced in full in a routine manner with amplification primers taken from exon number 5 and the flanking sequence information disclosed herein.

The information set forth in SEQ ID NO:1 and SEQ ID NO:3 was obtained as follows:

The first genomic clone "D" which contains exon 6, the longest exon, was isolated with the full length bovine A₁ adenosine receptor cDNA as a probe. See GenBank™/EMBL Data Bank Accession Number M86261; M. Olah et al., *supra*. A KpnI fragment of the genomic clone was subcloned into pGEM4Z (Promega) and sequenced with Sequenase version 2 DNA sequencing kit (United States Biochemical).

The second genomic clone "B" which contains exon 2-4 was isolated with a PCR fragment (based on bovine A₁AR cDNA sequence 526-854) as a probe. An EcoRI fragment (~ 7 kb) from the genomic clone restriction digest was subcloned and sequenced.

The genomic clone "A" which contains exon 1 was isolated with the EcoRI/NheI fragment (~ 110 bp) of human A₁AR cDNA clone 7A (see below) as a probe. A SphI fragment of this clone was subcloned.

The genomic clone "C" which contains exon 5 was isolated with a 50 mer synthetic oligonucleotide (based on human A₁AR cDNA sequence 755-804) as a probe. A KpnI fragment from this clone was subcloned.

cDNA Cloning. Human A₁AR cDNA clones were isolated from human brain (hippocampus) cDNA library in lambda ZAP II (Stratagene) with the SalI/HindIII fragment of the subclone of genomic clone "D" as a probe. The conditions used for library screening were the same as that used for genomic library screening. From ~ 1.0 x 10⁶

-24-

plaques, 56 positive clones were identified. Among nine analyzed clones, two containing longest inserts, 6A (2.61 kb) and 7A (2.48 kb), were sequenced. Clone 6A has a short 5'-untranslated sequence (120 bp), a full coding region (981 bp) and a complete 3'-untranslated sequence with polyadenylation signal and a poly (A) tail (1.51 kb). Clone 7A has a 410 bp 5'-untranslated sequence but missing a part of 3' sequence (1.09 kb rather than 1.51 kb). A more complete cDNA sequence of 2.9 kb, given herein as SEQ ID NO:5, is deduced from the combination of 6A (291-2900) and 7A (1-2480) sequences. The amino acid sequence coded for by the cDNA of SEQ ID NO:5 is set forth in SEQ ID NO:6.

Expression Vector Construction. An EcoRI/XbaI fragment of the human A₁AR cDNA clone 6A (291-1630) was subcloned into the pCMV5 expression vector in accordance with known techniques. See, e.g., M. Olah et al., *J. Biol. Chem.* 267, 10764-10770 (1992).

II. RESULTS

Screening of the human genomic library revealed multiple positive clones. Subcloning and sequencing of a number of these positive clones revealed that the human gene contained intronic interruptions of coding sequence based on the homology predicted from the known sequence of the bovine A₁ adenosine receptor cDNA. See M. Olah et al., *J. Biol. Chem.* 267, 10764-10770 (1992). Thus, unlike most G-protein coupled seven transmembrane domain receptors, the gene for the human A₁ adenosine receptor is not an intronless gene, but as can be seen in the schematic in Figure 1 (T. Bonner et al., *Science* 237, 527-532 (1987)) there are six exons interrupted by introns of various sizes. The full sequence of the exon structure and partial sequence of the intron structures are demonstrated in SEQ ID NO:1 and SEQ ID NO:3.

We next identified a human A₁ receptor cDNA by isolating a fragment from a human brain cDNA library. Two almost full length cDNAs were isolated. Clone 6A contains

a short 5' untranslated sequence, followed by a full coding sequence and the complete 3' untranslated sequence with a polyadenylation signal and a poly-A tail. Clone 7A has a 400-base pair 5' untranslated region but is missing a part of the 3' sequence as defined from the 6A clone. Thus, a more complete cDNA sequence can be deduced from a combination of the 6A and 7A sequences as given in SEQ ID NO:5. A direct comparison of the translated cDNAs from the bovine, dog, rat and human A₁ adenosine receptors (data not shown) indicates homology in the range of 90% between all the A₁ receptors. It should be noted that these sequence differences are important as can be verified by the fact that each receptor has a different pharmacology. See M. Olah et al., *supra*. An EcoRI/XbaI restriction fragment from the cDNA has been subcloned into the PCMV-5 expression vector as described in M. Olah et al., *supra*, and studies are currently underway expressing this receptor. A predicted structure for the human A₁AR is shown in Figure 2.

Mutagenesis studies carried out in the bovine A₁ adenosine receptor have revealed that mutation of Hist-278 to Leu-278 (found in transmembrane domain 7) dramatically decreases both agonist and antagonist binding by > 90%. This confirms previous biochemical data suggesting that histine residues are important for the binding of adenosine and its analogues to this receptor. In contrast, mutation of Hist-251 in transmembrane domain 6 to Leu-251 decreased antagonist affinity by four fold but had no effect on the affinity of agonists for the receptor. See M. Olah et al., *supra*. This suggests that one can manipulate the receptor in a way to alter how agonists and antagonists bind to the receptor, thereby providing potential therapeutic implications as described herein.

Mutations of phenylalanines 185 and 186 residues to leucines 185 and 186 in transmembrane domain-5 of the receptor results in a mutant receptor which binds agonists with high affinity, indicating that the coupling of the receptor to its G-protein is normal, but these receptors

-26-

fail to inhibit adenylyl cyclase in transfected CHO cells. This work was performed using the bovine A₁AR but the human A₁AR is identical in the region mutated (the hydrophobic pocket in the fifth transmembrane domain). This provides
5 further evidence that the receptors can be manipulated to make the receptor become essentially an antagonist receptor in that it will bind agonist with full high affinity but is incapable of activating or inhibiting an effector. While
10 receptor DNAs mutated in this manner are preferably human, they could be of other species (e.g., bovine, rat). These also has therapeutic potentials as described herein.

The foregoing examples are illustrative of the present invention, and are not to be construed as limiting thereof. The invention is defined by the following claims,
15 with equivalents of the claims to be included therein.

-27-

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Stiles, Gary L.
Ren, Hongzu
Olah, Mark E.

(ii) TITLE OF INVENTION: DNA Encoding the Human A1 Adenosine Receptor

(iii) NUMBER OF SEQUENCES: 6

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(F) ZIP: 28234

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Sibley, Kenneth D.
(B) REGISTRATION NUMBER: 31,665

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 919-881-3140
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(C) TELEX: 575102

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1513 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

-28-

(A) NAME/KEY: intron
(B) LOCATION: 1..71

(ix) FEATURE:
(A) NAME/KEY: exon
(B) LOCATION: 72..161

(ix) FEATURE:
(A) NAME/KEY: intron
(B) LOCATION: 162..525

(ix) FEATURE:
(A) NAME/KEY: exon
(B) LOCATION: 526..679

(ix) FEATURE:
(A) NAME/KEY: intron
(B) LOCATION: 680..1030

(ix) FEATURE:
(A) NAME/KEY: exon
(B) LOCATION: 1031..1428

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1088..1426

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CAGGCGCCCG AGTCGAGTCC CAGCCAGCTA CCATCCCTCT GGAGCTTACC GGCCGGCCTT	60
GGCTTCCCCA GGAATCCCTG GAGCTAGCGG CTGCTGAAGG CGTCGAGGTG TGGGGGCACT	120
TGGACAGAAC AGTCAGGCAG CCGGGAGCTC TGCCAGCTTT GGTGACCTTG GGTAAGTCTG	180
AGTCTCGGTT CACCCCTGGG GCTCCCCAAT GGGGGTGAG GAGAGGGTTG AAAGAAGAGA	240
CCGGAATGGC CCCTTGGGGC AGGCCATGGG CAAGGTTCCC CGACAGAGCT GGAACGGGAC	300
CAGAGGACTG CTAAGATCCA GGCACCAGGA CGGGTCTCAA GGTGGGTGGG CGCAGGGCAG	360
GTGCGGGCAC GCTGAGGGAA TAGGGAGAAA ACGCCCCAGC CTTGTCCTGG GCTCCGTCCC	420
CAGACCCACG TCTGCCACCC CAGTCCCAGG TCGAAACAG GGGGCGCTAC CTCTTTAAAA	480
GCGTCCGGGG CTGAGTCTCT GCCGTACCAT GTGATTGCTT GAAAGGCCGG GCTGGGAGCG	540
CTGCGGCGGG AGCCGGAGGA CTATGAGCTG CCGCGCGTTG TCCAGAGCCC AGCCAGCCCC	600
TACGCGCGCG GCCCGGAGCT CTGTTCCCTG GAACTTTGGG CACTGCCTCT GGGACCCCTG	660
CCGGCCAGCA GGCAGGATGG TGAGTCCCT GCATCCTGTT CTGTGCACAG GGGTGGGCAG	720
AGCCAGTCAT GGGAGACCCC TCTGTGCGTG TGTCTGTGTG TGCGCGCGCG CTGGGAGCTG	780

-29-

CCTCACACCT CATAAAAAAG CCAGTGGAGG AGTGAGGCTG CTATTTTAAG TTGCTGAATG	840
GAACCTCTGG GAATGATAAA GGAAAGGGAC AAAGATTAGG CAGAGAAGGG TCCGGGTGCC	900
CCTCCAGCCT GGGTAGGAGC TGCATGTGAC AAGTGGGACA CATCACAGGG TACCTGGAGT	960
TCCAGGGCAG CCTGAGCTCC CTGCCCCTCC CAGACCGGTC TCCCCATCCC AGGCTTCCT	1020
GACCACACAG GTGCTTGCTT CGTGCCCTT GGTGCCGTC TGCTGATGTG CCCAGCCTGT	1080
GCCCGCC ATG CCG CCC TCC ATC TCA GCT TTC CAG GCC GCC TAC ATC GGC	1129
Met Pro Pro Ser Ile Ser Ala Phe Gln Ala Ala Tyr Ile Gly	
1 5 10	
ATC GAG GTG CTC ATC GCC CTG GTC TCT GTG CCC GGG AAC GTG CTG GTG	1177
Ile Glu Val Leu Ile Ala Leu Val Ser Val Pro Gly Asn Val Leu Val	
15 20 25 30	
ATC TGG GCG GTG AAG GTG AAC CAG GCG CTG CGG GAT GCC ACC TTC TGC	1225
Ile Trp Ala Val Lys Val Asn Gln Ala Leu Arg Asp Ala Thr Phe Cys	
35 40 45	
TTC ATC GTG TCG CTG GCG GTG GCT GAT GTG GCC GTG GGT GCC CTG GTC	1273
Phe Ile Val Ser Leu Ala Val Ala Asp Val Ala Val Gly Ala Leu Val	
50 55 60	
ATC CCC CTC GCC ATC CTC ATC AAC ATT GGG CCA CAG ACC TAC TTC CAC	1321
Ile Pro Leu Ala Ile Leu Ile Asn Ile Gly Pro Gln Thr Tyr Phe His	
65 70 75	
ACC TGC CTC ATG GTT GCC TGT CCG GTC CTC ATC CTC ACT CAG AGC TCC	1369
Thr Cys Leu Met Val Ala Cys Pro Val Leu Ile Leu Thr Gln Ser Ser	
80 85 90	
ATC CTG GCC CTG CTG GCA ATT GCT GTG GAC CGC TAC CTC CGG GTC AAG	1417
Ile Leu Ala Leu Leu Ala Ile Ala Val Asp Arg Tyr Leu Arg Val Lys	
95 100 105 110	
ATC CCT CTC CGGTGAGTCC ACAGCGCCGA AGTACTCGCA GCACCACATG	1466
Ile Pro Leu	
ATGGCTGGCT TGAGGGCCAT CTAGAAAGGA AAAAAGGTAG AGCATAA	1513

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 113 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

-30-

Met Pro Pro Ser Ile Ser Ala Phe Gln Ala Ala Tyr Ile Gly Ile Glu
 1 5 10 15
 Val Leu Ile Ala Leu Val Ser Val Pro Gly Asn Val Leu Val Ile Trp
 20 25 30
 Ala Val Lys Val Asn Gln Ala Leu Arg Asp Ala Thr Phe Cys Phe Ile
 35 40 45
 Val Ser Leu Ala Val Ala Asp Val Ala Val Gly Ala Leu Val Ile Pro
 50 55 60
 Leu Ala Ile Leu Ile Asn Ile Gly Pro Gln Thr Tyr Phe His Thr Cys
 65 70 75 80
 Leu Met Val Ala Cys Pro Val Leu Ile Leu Thr Gln Ser Ser Ile Leu
 85 90 95
 Ala Leu Leu Ala Ile Ala Val Asp Arg Tyr Leu Arg Val Lys Ile Pro
 100 105 110
 Leu

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2180 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: intron
- (B) LOCATION: 1..29

(ix) FEATURE:

- (A) NAME/KEY: exon
- (B) LOCATION: 30..2125

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 32..616

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

TTGACCTGCA GGTGACCTG CAGGTCAACG G ATC CTC TCC TTC GTG GTG GGA 52
 Ile Leu Ser Phe Val Val Gly
 1 5
 CTG ACC CCT ATG TTT GGC TGG AAC AAT CTG AGT GCG GTG GAG CGG GCC 100
 Leu Thr Pro Met Phe Gly Trp Asn Asn Leu Ser Ala Val Glu Arg Ala

-31-

10	15	20	
TGG GCA GCC AAC GGC AGC ATG GGG GAG CCC GTG ATC AAG TGC GAG TTC Trp Ala Ala Asn Gly Ser Met Gly Glu Pro Val Ile Lys Cys Glu Phe 25 30 35			148
GAG AAG GTC ATC AGC ATG GAG TAC ATG GTC TAC TTC AAC TTC TTT GTG Glu Lys Val Ile Ser Met Glu Tyr Met Val Tyr Phe Asn Phe Phe Val 40 45 50 55			196
TGG GTG CTG CCC CCG CTT CTC CTC ATG GTC CTC ATC TAC CTG GAG GTC Trp Val Leu Pro Pro Leu Leu Leu Met Val Leu Ile Tyr Leu Glu Val 60 65 70			244
TTC TAC CTA ATC CGC AAG CAG CTC AAC AAG AAG GTG TCG GCC TCC TCC Phe Tyr Leu Ile Arg Lys Gln Leu Asn Lys Lys Val Ser Ala Ser Ser 75 80 85			292
GGC GAC CCG CAG AAG TAC TAT GGG AAG GAG CTG AAG ATC GCC AAG TCG Gly Asp Pro Gln Lys Tyr Tyr Gly Lys Glu Leu Lys Ile Ala Lys Ser 90 95 100			340
CTG GCC CTC ATC CTC TTC CTC TTT GCC CTC AGC TGG CTG CCT TTG CAC Leu Ala Leu Ile Leu Phe Leu Phe Ala Leu Ser Trp Leu Pro Leu His 105 110 115			388
ATC CTC AAC TGC ATC ACC CTC TTC TGC CCG TCC TGC CAC AAG CCC AGC Ile Leu Asn Cys Ile Thr Leu Phe Cys Pro Ser Cys His Lys Pro Ser 120 125 130 135			436
ATC CTT ACC TAC ATT GCC ATC TTC CTC ACG CAC GGC AAC TCG GCC ATG Ile Leu Thr Tyr Ile Ala Ile Phe Leu Thr His Gly Asn Ser Ala Met 140 145 150			484
AAC CCC ATT GTC TAT GCC TTC CGC ATC CAG AAG TTC CGC GTC ACC TTC Asn Pro Ile Val Tyr Ala Phe Arg Ile Gln Lys Phe Arg Val Thr Phe 155 160 165			532
CTT AAG ATT TGG AAT GAC CAT TTC CGC TGC CAG CCT GCA CCT CCC ATT Leu Lys Ile Trp Asn Asp His Phe Arg Cys Gln Pro Ala Pro Pro Ile 170 175 180			580
GAC GAG GAT CTC CCA GAA GAG AGG CCT GAT GAC TAGACCCCGC CTTCGCTCC Asp Glu Asp Leu Pro Glu Glu Arg Pro Asp Asp 185 190 195			633
CACCAGCCCA CATCCAGTGG GGTCTCAGTC CAGTCCTCAC ATGCCCGCTG TCCCAGGGGT			693
CTCCCTGAGC CTGCCCCAGC TGGGCTGTTG GCTGGGGGCA TGGGGGAGGC TCTGAAGAGA			753
TACCCACAGA GTGTGGTCCC TCCACTAGGA GTTAACTACC CTACACCTCT GGGCCCTGCA			813
GGAGGCCTGG GAGGGCAAGG GTCCTACGGA GGGACCAGGT GTCTAGAGGC AACAGTGTTC			873
TGAGCCCCCA CCTGCCTGAC CATCCCATGA GCAGTCCAGC GCTTCAGGGC TGGGCAGGTC			933

-32-

CTGGGGAGGC TGAGACTGCA GAGGAGCCAC CTGGGCTGGG AGAAGGTGCT TGGGCTTCTG 993
 CGGTGAGGCA GGGGAGTCTG CTTGTCTTAG ATGTTGGTGG TGCAGCCCCA GGACCAAGCT 1053
 TAAGGAGAGG AGAGCATCTG CTCTGAGACG GATGGAAGGA GAGAGGTTGA GGATGCACTG 1113
 GCCTGTTCTG TAGGAGAGAC TGGCCAGAGG CAGCTAAGGG GCAGGAATCA AGGAGCCTCC 1173
 GTTCCACCT CTGAGGACTC TGGACCCAG GCCATACCAG GTGCTAGGGT GCCTGCTCTC 1233
 CTTGCCCTGG GCCAGCCAG GATTGTACGT GGGAGAGGCA GAAAGGGTAG GTTCAGTAAT 1293
 CATTCTGAT GATTGCTGG AGTGCTGGCT CCACGCCCTG GGGAGTGAGC TTGGTGCGGT 1353
 AGGTGCTGGC CTCAACAGC CACGAGGTGG TAGCTCTGAG CCCTCCTTCT TGCCCTGAGC 1413
 TTTCCGGGGA GGAGCCTGGA GTGTAATTAC CTGTCATCTG GGCCACCAGC TCCACTGGCC 1473
 CCCGTTGCCG GGCCTGGACT GTCCTAGGTG ACCCATCTC TGCTGCTTCT GGGCCTGATG 1533
 GAGAGGAGAA CACTAGACAT GCCAACTCGG GAGCATTCTG CCTGCCTGGG AACGGGGTGG 1593
 ACGAGGGAGT GTCTGTAAGG ACTCAGTGT GACTGTAGGC GCCCCTGGGG TGGGTTTAGC 1653
 AGGCTGCAGC AGGCAGAGGA GGAGTACCCC CCTGAGAGCA TGTGGGGGAA GGCCTTGCTG 1713
 TCATGTGAAT CCCTCAATAC CCCTAGTATC TGGCTGGGT TTCAGGGGCT TTGGAAGCTC 1773
 TGTTGCAGGT GTCCGGGGGT CTAGGACTTT AGGGATCTGG GATCTGGGGA AGGACCAACC 1833
 CATGCCCTGC CAAGCCTGGA GCCCCTGTGT TGGGGGGCAA GGTGGGGGAG CCTGGAGCCC 1893
 CTGTGTGGGA GGGCGAGGCG GGGGAGCCTG GAGCCCCTGT GTGGGAGGGC GAGGCGGGGG 1953
 ATCCTGGAGC CCCTGTGTCG GGGGGCGAGG GAGGGGAGGT GGCCGTCGGT TGACCTTCTG 2013
 AACATGAGTG TCAACTCCAG GACTTGCTTC CAAGCCCTTC CCTCTGTTGG AAATTGGGTG 2073
 TGCCCTGGCT CCCAAGGGAG GCCCATGTGA CTAATAAAAA ACTGTGAACC CTGTGGAGAG 2133
 CACATTGCTG GGCGCCCATC CCCACCACTG TTGAGGGCAT GAAGACA 2180

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 194 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ile Leu Ser Phe Val Val Gly Leu Thr Pro Met Phe Gly Trp Asn Asn
 1 5 10 15

-33-

Leu Ser Ala Val Glu Arg Ala Trp Ala Ala Asn Gly Ser Met Gly Glu
 20 25 30
 Pro Val Ile Lys Cys Glu Phe Glu Lys Val Ile Ser Met Glu Tyr Met
 35 40 45
 Val Tyr Phe Asn Phe Phe Val Trp Val Leu Pro Pro Leu Leu Leu Met
 50 55 60
 Val Leu Ile Tyr Leu Glu Val Phe Tyr Leu Ile Arg Lys Gln Leu Asn
 65 70 75 80
 Lys Lys Val Ser Ala Ser Ser Gly Asp Pro Gln Lys Tyr Tyr Gly Lys
 85 90 95
 Glu Leu Lys Ile Ala Lys Ser Leu Ala Leu Ile Leu Phe Leu Phe Ala
 100 105 110
 Leu Ser Trp Leu Pro Leu His Ile Leu Asn Cys Ile Thr Leu Phe Cys
 115 120 125
 Pro Ser Cys His Lys Pro Ser Ile Leu Thr Tyr Ile Ala Ile Phe Leu
 130 135 140
 Thr His Gly Asn Ser Ala Met Asn Pro Ile Val Tyr Ala Phe Arg Ile
 145 150 155 160
 Gln Lys Phe Arg Val Thr Phe Leu Lys Ile Trp Asn Asp His Phe Arg
 165 170 175
 Cys Gln Pro Ala Pro Pro Ile Asp Glu Asp Leu Pro Glu Glu Arg Pro
 180 185 190
 Asp Asp

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2900 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 411..1391

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATGAGTGTCA GAAGTGTGAA GGGTGCCTGT TCTGAATCCC AGAGCCTCCT CTCCTCTGT

-34-

GAGGCTGGCA GGTGAGGAAG GGTTTAACCT CACTGGAAGG AATCCCTGGA GCTAGCGGCT	120
GCTGAAGGCG TCGAGGTGTG GGGGCACTTG GACAGAACAG TCAGGCAGCC GGGAGCTCTG	180
CCAGCTTTGG TGACCTTGGG CCGGGCTGGG AGCGCTGCGG CGGGAGCCGG AGGACTATGA	240
GCTGCCGCGC GTTGTCCAGA GCCCAGCCCA GCCCTACGCG CGCGGCCCGG AGCTCTGTTC	300
CCTGGAACCT TGGGCACTGC CTCTGGGACC CCTGCCGGCC AGCAGGCAGG ATGGTGCTTG	360
CCTCGTGCCC CTTGGTGCCC GTCTGCTGAT GTGCCAGCC TGTGCCCGCC ATG CCG	416
Met Pro	
1	
CCC TCC ATC TCA GCT TTC CAG GCC GCC TAC ATC GGC ATC GAG GTG CTC	464
Pro Ser Ile Ser Ala Phe Gln Ala Ala Tyr Ile Gly Ile Glu Val Leu	
5 10 15	
ATC GCC CTG GTC TCT GTG CCC GGG AAC GTG CTG GTG ATC TGG GCG GTG	512
Ile Ala Leu Val Ser Val Pro Gly Asn Val Leu Val Ile Trp Ala Val	
20 25 30	
AAG GTG AAC CAG GCG CTG CGG GAT GCC ACC TTC TGC TTC ATC GTG TCG	560
Lys Val Asn Gln Ala Leu Arg Asp Ala Thr Phe Cys Phe Ile Val Ser	
35 40 45 50	
CTG GCG GTG GCT GAT GTG GCC GTG GGT GCC CTG GTC ATC CCC CTC GCC	608
Leu Ala Val Ala Asp Val Ala Val Gly Ala Leu Val Ile Pro Leu Ala	
55 60 65	
ATC CTC ATC AAC ATT GGG CCA CAG ACC TAC TTC CAC ACC TGC CTC ATG	656
Ile Leu Ile Asn Ile Gly Pro Gln Thr Tyr Phe His Thr Cys Leu Met	
70 75 80	
GTT GCC TGT CCG GTC CTC ATC CTC ACC CAG AGC TCC ATC CTG GCC CTG	704
Val Ala Cys Pro Val Leu Ile Leu Thr Gln Ser Ser Ile Leu Ala Leu	
85 90 95	
CTG GCA ATT GCT GTG GAC CGC TAC CTC CGG GTC AAG ATC CCT CTC CGG	752
Leu Ala Ile Ala Val Asp Arg Tyr Leu Arg Val Lys Ile Pro Leu Arg	
100 105 110	
TAC AAG ATG GTG GTG ACC CCC CGG AGG GCG GCG GTG GCC ATA GCC GGC	800
Tyr Lys Met Val Val Thr Pro Arg Arg Ala Ala Val Ala Ile Ala Gly	
115 120 125 130	
TGC TGG ATC CTC TCC TTC GTG GTG GGA CTG ACC CCT ATG TTT GGC TGG	848
Cys Trp Ile Leu Ser Phe Val Val Gly Leu Thr Pro Met Phe Gly Trp	
135 140 145	

-35-

AAC AAT CTG AGT GCG GTG GAG CGG GCC TGG GCA GCC AAC GGC AGC ATG Asn Asn Leu Ser Ala Val Glu Arg Ala Trp Ala Ala Asn Gly Ser Met 150 155 160	896
GGG GAG CCC GTG ATC AAG TGC GAG TTC GAG AAG GTC ATC AGC ATG GAG Gly Glu Pro Val Ile Lys Cys Glu Phe Glu Lys Val Ile Ser Met Glu 165 170 175	944
TAC ATG GTC TAC TTC AAC TTC TTT GTG TGG GTG CTG CCC CCG CTT CTC Tyr Met Val Tyr Phe Asn Phe Phe Val Trp Val Leu Pro Pro Leu Leu 180 185 190	992
CTC ATG GTC CTC ATC TAC CTG GAG GTC TTC TAC CTA ATC CGC AAG CAG Leu Met Val Leu Ile Tyr Leu Glu Val Phe Tyr Leu Ile Arg Lys Gln 195 200 205 210	1040
CTC AAC AAG AAG GTG TCG GCC TCC TCC GGC GAC CCG CAG AAG TAC TAT Leu Asn Lys Lys Val Ser Ala Ser Ser Gly Asp Pro Gln Lys Tyr Tyr 215 220 225	1088
GGG AAG GAG CTG AAG ATC GCC AAG TCG CTG GCC CTC ATC CTC TTC CTC Gly Lys Glu Leu Lys Ile Ala Lys Ser Leu Ala Leu Ile Leu Phe Leu 230 235 240	1136
TTT GCC CTC AGC TGG CTG CCT TTG CAC ATC CTC AAC TGC ATC ACC CTC Phe Ala Leu Ser Trp Leu Pro Leu His Ile Leu Asn Cys Ile Thr Leu 245 250 255	1184
TTC TGC CCG TCC TGC CAC AAG CCC AGC ATC CTT ACC TAC ATT GCC ATC Phe Cys Pro Ser Cys His Lys Pro Ser Ser Ile Leu Thr Tyr Ile Ala Ile 260 265 270	1232
TTC CTC ACG CAC GGC AAC TCG GCC ATG AAC CCC ATT GTC TAT GCC TTC Phe Leu Thr His Gly Asn Ser Ala Met Asn Pro Ile Val Tyr Ala Phe 275 280 285 290	1280
CGC ATC CAG AAG TTC CGC GTC ACC TTC CTT AAG ATT TGG AAT GAC CAT Arg Ile Gln Lys Phe Arg Val Thr Phe Leu Lys Ile Trp Asn Asp His 295 300 305	1328
TTC CGC TGC CAG CCT GCA CCT CCC ATT GAC GAG GAT CTC CCA GAA GAG Phe Arg Cys Gln Pro Ala Pro Pro Ile Asp Glu Asp Leu Pro Glu Glu 310 315 320	1376
AGG CCT GAT GAC TAGACCCCGC CTTCGCTCC CACCAGCCCA CATCCAGTGG Arg Pro Asp Asp 325	1428
GGTCTCAGTC CAGTCCTCAC ATGCCCCTG TCCAGGGGT CTCCCTGAGC CTGCCCCAGC	1488
TGGGCTGTTG GCTGGGGGCA TGGGGGAGGC TCTGAAGAGA TACCCACAGA GTGTGGTCCC	1548
TCCACTAGGA GTTAACTACC CTACACCTCT GGGCCCTGCA GGAGGCCTGG GAGGGCAAGG	1608
GTCCTACGGA GGGACCAGGT GTCTAGAGGC AACAGTGTTC TGAGCCCCCA CCTGCCTGAC	1668

-36-

CATCCCATGA GCAGTCCAGC GCTTCAGGGC TGGGCAGGTC CTGGGGAGGC TGAGACTGCA	1728
GAGGAGCCAC CTGGGCTGGG AGAAGGTGCT TGGGCTTCTG CGGTGAGGCA GGGGAGTCTG	1788
CTTGTCTTAG ATGTTGGTGG TGCAGCCCCA GGACCAAGCT TAAGGAGAGG AGAGCATCTG	1848
CTCTGAGACG GATGGAAGGA GAGAGGTTGA GGATGCACTG GCCTGTTCTG TAGGAGAGAC	1908
TGGCCAGAGG CAGCTAAGGG GCAGGAATCA AGGAGCCTCC GTTCCCACCT CTGAGGACTC	1968
TGGACCCAG GCCATACCAG GTGCTAGGGT GCCTGCTCTC CTTGCCCTGG GCCAGCCAG	2028
GATTGTACGT GGGAGAGGCA GAAAGGGTAG GTTCAGTAAT CATTTCTGAT GATTTGCTGG	2088
AGTGCTGGCT CCACGCCCTG GGGAGTGAGC TTGGTGCGGT AGGTGCTGGC CTCAAACAGC	2148
CACGAGGTGG TAGCTCTGAG CCCTCCTTCT TGCCCTGAGC TTTCCGGGGA GGAGCCTGGA	2208
GTGTAATTAC CTGTCATCTG GGCCACCAGC TCCACTGGCC CCCGTTGCCG GGCCTGGACT	2268
GTCCTAGGTG ACCCCATCTC TGCTGCTTCT GGGCCTGATG GAGAGGAGAA CACTAGACAT	2328
GCCAACTCGG GAGCATTCTG CCTGCCTGGG AACGGGGTGG ACGAGGGAGT GTCTGTAAGG	2388
ACTCAGTGTT GACTGTAGGC GCCCCTGGGG TGGGTTTAGC AGGCTGCAGC AGGCAGAGGA	2448
GGAGTACCCC CCTGAGAGCA TGTGGGGGAA GGCCTTGCTG TCATGTGAAT CCCTCAATAC	2508
CCCTAGTATC TGGCTGGGTT TTCAGGGGCT TTGGAAGCTC TGTTCAGGT GTCCGGGGGT	2568
CTAGGACTTT AGGGATCTGG GATCTGGGGA AGGACCAACC CATGCCCTGC CAAGCCTGGA	2628
GCCCCTGTGT TGGGGGGCAA GGTGGGGGAG CCTGGAGCCC CTGTGTGGGA GGGCGAGGCG	2688
GGGGAGCCTG GAGCCCCTGT GTGGGAGGGC GAGGCGGGGG ATCCTGGAGC CCCTGTGTCTG	2748
GGGGGCGAGG GAGGGGAGGT GGCCGTCGGT TGACCTTCTG AACATGAGTG TCAACTCCAG	2808
GACTTGCTTC CAAGCCCTTC CCTCTGTTGG AAATTGGGTG TGCCCTGGCT CCCAAGGGAG	2868
GCCCATGTGA CTAATAAAAA ACTGTGAACC CT	2900

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 326 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

-37-

Met Pro Pro Ser Ile Ser Ala Phe Gln Ala Ala Tyr Ile Gly Ile Glu
 1 5 10 15
 Val Leu Ile Ala Leu Val Ser Val Pro Gly Asn Val Leu Val Ile Trp
 20 25 30
 Ala Val Lys Val Asn Gln Ala Leu Arg Asp Ala Thr Phe Cys Phe Ile
 35 40 45
 Val Ser Leu Ala Val Ala Asp Val Ala Val Gly Ala Leu Val Ile Pro
 50 55 60
 Leu Ala Ile Leu Ile Asn Ile Gly Pro Gln Thr Tyr Phe His Thr Cys
 65 70 75 80
 Leu Met Val Ala Cys Pro Val Leu Ile Leu Thr Gln Ser Ser Ile Leu
 85 90 95
 Ala Leu Leu Ala Ile Ala Val Asp Arg Tyr Leu Arg Val Lys Ile Pro
 100 105 110
 Leu Arg Tyr Lys Met Val Val Thr Pro Arg Arg Ala Ala Val Ala Ile
 115 120 125
 Ala Gly Cys Trp Ile Leu Ser Phe Val Val Gly Leu Thr Pro Met Phe
 130 135 140
 Gly Trp Asn Asn Leu Ser Ala Val Glu Arg Ala Trp Ala Ala Asn Gly
 145 150 155 160
 Ser Met Gly Glu Pro Val Ile Lys Cys Glu Phe Glu Lys Val Ile Ser
 165 170 175
 Met Glu Tyr Met Val Tyr Phe Asn Phe Phe Val Trp Val Leu Pro Pro
 180 185 190
 Leu Leu Leu Met Val Leu Ile Tyr Leu Glu Val Phe Tyr Leu Ile Arg
 195 200 205
 Lys Gln Leu Asn Lys Lys Val Ser Ala Ser Ser Gly Asp Pro Gln Lys
 210 215 220
 Tyr Tyr Gly Lys Glu Leu Lys Ile Ala Lys Ser Leu Ala Leu Ile Leu
 225 230 235 240
 Phe Leu Phe Ala Leu Ser Trp Leu Pro Leu His Ile Leu Asn Cys Ile
 245 250 255
 Thr Leu Phe Cys Pro Ser Cys His Lys Pro Ser Ile Leu Thr Tyr Ile
 260 265 270
 Ala Ile Phe Leu Thr His Gly Asn Ser Ala Met Asn Pro Ile Val Tyr
 275 280 285

-38-

Ala Phe Arg Ile Gln Lys Phe Arg Val Thr Phe Leu Lys Ile Trp Asn
290 295 300

Asp His Phe Arg Cys Gln Pro Ala Pro Pro Ile Asp Glu Asp Leu Pro
305 310 315 320

Glu Glu Arg Pro Asp Asp
325

THAT WHICH IS CLAIMED IS:

1. Isolated DNA encoding an A_1 adenosine receptor selected from the group consisting of:

(a) isolated human genomic DNA consisting essentially of DNA which encodes the human A_1 adenosine receptor given herein as SEQ ID NO:6 and which contains the DNA sequences given herein as SEQ ID NO:1 and SEQ ID NO:3;

(b) isolated human genomic DNA which hybridizes to isolated DNA of (a) above under conditions represented by a wash stringency of 0.3M NaCl, 0.03M sodium citrate, and 0.1% SDS at 60°C, which is at least 65% homologous to isolated DNA of (a) above, and which encodes an A_1 adenosine receptor; and

(c) isolated DNA differing from the isolated DNAs of (a) and (b) above in nucleotide sequence due to the degeneracy of the genetic code, and which encodes an A_1 adenosine receptor.

2. Isolated DNA according to claim 1 consisting essentially of isolated human genomic DNA which encodes the human A_1 adenosine receptor given herein as SEQ ID NO:6 and which contains the DNA sequences given herein as SEQ ID NO:1 and SEQ ID NO:3.

3. A recombinant DNA sequence comprising vector DNA and a DNA according to claim 1.

4. A recombinant DNA sequence according to claim 3, wherein said vector DNA comprises a vector selected from the group consisting of plasmids, adenoviruses, and cytomegaloviruses.

-40-

5. A recombinant DNA sequence according to claim 3, wherein said vector DNA comprises a baculovirus vector.

6. A host cell containing a recombinant DNA sequence of claim 3 and capable of expressing the encoded protein.

7. A host cell according to claim 6, wherein said host cell is a mammalian cell.

8. A host cell according to claim 6, wherein said host cell is selected from the group consisting of baby hamster kidney cells, mouse cells, human embryo cells, and chinese hamster ovary cells.

9. A host cell according to claim 6, wherein said host cell is an insect cell.

10. Isolated DNA encoding an A₁ adenosine receptor selected from the group consisting of:

(a) isolated DNA which encodes a human A₁ adenosine receptor consisting essentially of DNA having the sequence given herein as SEQ ID NO:5;

(b) isolated DNA which hybridizes to isolated DNA of (a) above under conditions represented by a wash stringency of 0.3M NaCl, 0.03M sodium citrate, and 0.1% SDS at 60°C, which is at least 93% homologous to isolated DNA of (a) above, and which encodes a human A₁ adenosine receptor; and

(c) isolated DNA differing from the isolated DNAs of (a) and (b) above in nucleotide sequence due to the degeneracy of the genetic code, and which encodes a human A₁ adenosine receptor.

-41-

11. Isolated DNA according to claim 10 which encodes a human A₁ adenosine receptor consisting essentially of DNA having the sequence given herein as SEQ ID NO:5.

12. A recombinant DNA sequence comprising vector DNA and a DNA according to claim 10.

13. A recombinant DNA sequence according to claim 12, wherein said vector DNA comprises a vector selected from the group consisting of plasmids, adenoviruses, and cytomegaloviruses.

14. A recombinant DNA sequence according to claim 12, wherein said vector DNA comprises a baculovirus vector.

15. A host cell containing a recombinant DNA sequence of claim 12 and capable of expressing the encoded protein.

16. A host cell according to claim 15, wherein said host cell is a mammalian cell.

17. A host cell according to claim 15, wherein said host cell is selected from the group consisting of baby hamster kidney cells, mouse cells, human embryo cells, and chinese hamster ovary cells.

18. A host cell according to claim 15, wherein said host cell is an insect cell.

19. Oligonucleotides capable of binding to the introns of the A₁ adenosine receptor, said oligonucleotides selected from the group consisting of:

(a) (i) isolated DNA consisting essentially of the first intron of the human genomic DNA encoding the human A₁ adenosine receptor according to SEQ ID NO:6, (ii) isolated DNA consisting essentially of the second intron of the human genomic DNA encoding the human A₁ adenosine receptor according to SEQ ID NO:6, (iii) isolated DNA consisting essentially of the third intron of the human genomic DNA encoding the human A₁ adenosine receptor according to SEQ ID NO:6, (iv) isolated DNA consisting essentially of the fourth intron of the human genomic DNA encoding the human A₁ adenosine receptor according to SEQ ID NO:6, and (v) isolated DNA consisting essentially of the fifth intron of the human genomic DNA encoding the human A₁ adenosine receptor according to SEQ ID NO:6; and

(b) isolated DNA which hybridizes to the isolated DNAs of (a) above under conditions represented by a wash stringency of 0.3M NaCl, 0.03M sodium citrate, and 0.1% SDS at 60°C, and which is at least 75% homologous to isolated DNA of (a) above.

20. An oligonucleotide according to claim 19 labelled with a detectable group.

21. A recombinant DNA sequence comprising vector DNA and an oligonucleotide according to claim 19.

-43-

22. A recombinant DNA sequence according to claim 21, wherein said DNA sequence is capable of exchanging said oligonucleotide with a homologous nucleotide in a suitable cell by homologous recombination.

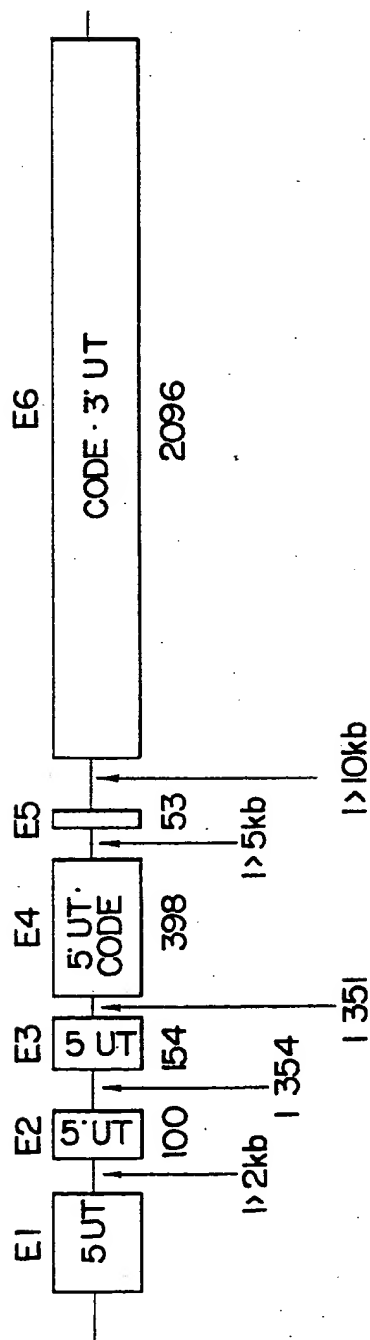


FIG. 1.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US93/06782

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :C12N 15/12, 15/63

US CL :435/252.3, 320.1; 536/23.5

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/69.1, 172.3, 252.3, 320.1; 530/350; 536/23.5

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, STN/MEDLINE

search terms: A1 adenosine receptor#, cDNA, DNA, clone#

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Annual Review of Entomology, Vol. 34, issued 1989, S. Maeda, "EXPRESSION OF FOREIGN GENES IN INSECTS USING BACULOVIRUS VECTORS", pages 351 to 372, see especially pages 364 and 365.	5, 9, 14, 18
Y	The EMBO Journal, Vol. 10, No. 7, issued 1991, Libert et.al., "The orphan receptor cDNA RDC7 encodes an A1 adenosine receptor", pages 1677 to 1682, see entire document.	1-22
Y	GENOMICS, Vol. 11, issued September 1991, Libert et.al., "Chromosomal Mapping of A1 and A2 Adenosine Receptors, VIP Receptor, and a New Subtype of Serotonin Receptor", pages 225 to 227, see entire document.	1-22

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be part of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Z" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

04 November 1993

Date of mailing of the international search report

NOV 18 1993

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/06782

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	NATURE, Vol. 342, issued 21 December 1989, Giros et.al., "Alternative splicing directs the expression of two D ₂ dopamine receptor isoforms", pages 923 to 926, see page 924.	1-22
Y	NATURE, Vol. 336, issued 22 December 1988, Bunzow et.al., "Cloning and expression of a rat D ₂ dopamine receptor cDNA", pages 783 to 787, see entire document.	1-22

